

# Functional Characterization of a Dehydratase Domain from the Pikromycin Polyketide Synthase

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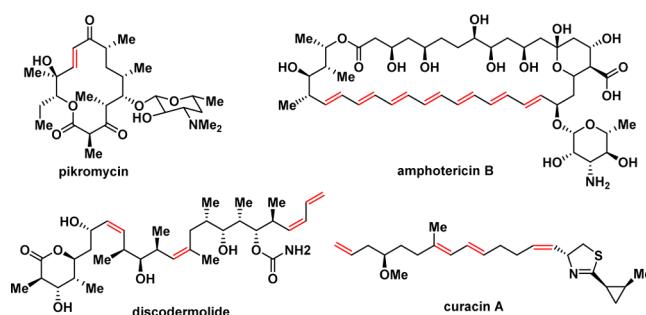
**S** Supporting Information

**ABSTRACT:** Metabolic engineering of polyketide synthase (PKS) pathways represents a promising approach to natural products discovery. The dehydratase (DH) domains of PKSs, which generate an  $\alpha,\beta$ -unsaturated bond through a dehydration reaction, have been poorly studied compared with other domains, likely because of the simple nature of the chemical reaction they catalyze and the lack of a convenient assay to measure substrate turnover. Herein we report the first steady-state kinetic analysis of a PKS DH domain employing LC–MS/MS analysis for product quantitation. PikDH2 was selected as a model DH domain. Its substrate specificity and mechanism were interrogated with a systematic series of synthetic triketide substrates containing a nonhydrolyzable thioether linkage as well as by site-directed mutagenesis, evaluation of the pH dependence of the catalytic efficiency ( $V_{\max}/K_M$ ), and kinetic characterization of a mechanism-based inhibitor. These studies revealed that PikDH2 converts D-alcohol substrates to *trans*-olefin products. The reaction is reversible with equilibrium constants ranging from 1.2 to 2. Moreover, the enzyme activity is robust, and PikDH2 was used on a preparative scale for the chemoenzymatic synthesis of unsaturated triketide products. PikDH2 was shown to possess remarkably strict substrate specificity and is unable to turn over substrates that are epimeric at the  $\beta$ -,  $\gamma$ -, or  $\delta$ -position. We also demonstrated that PikDH2 has a key ionizable group with a  $pK_a$  of 7.0 and can be irreversibly inactivated through covalent modification by a mechanism-based inhibitor, which provides a foundation for future structural studies to elucidate substrate–protein interactions.

Polyketides derived from modular type I polyketide synthases (PKSs) have drawn enormous attention and interest from chemists for decades because of their intricate structures, stereochemical complexity, and diverse pharmacological activities. Sharing the same evolutionary history with fatty acid synthases (FASs),<sup>1</sup> modular type I PKSs employ similar assembly-line molecular machinery wherein the chain intermediates remain covalently attached to acyl carrier protein (ACP) domains during biosynthesis. Unlike FASs, the presence of the three processing domains in PKSs, ketoreductase (KR), dehydratase (DH), and enoylreductase (ER), are varied in each module, leading to a fully reduced, partially reduced, or unreduced segment on the polyketide chain. Because of their

assembly-line attributes and varying combinations of processing domains, PKSs have been exploited through combinatorial biosynthesis and metabolic engineering to provide large libraries of polyketide analogues.<sup>2</sup>

Many polyketides, such as the archetypical macrolide antibiotic pikromycin, the antifungal polyene amphotericin B, the linear polyketide discodermolide, and the mixed non-ribosomal peptide–polyketide curacin, contain one or more double bonds that serve as conformational constraints and are essential for biological activity (Figure 1).<sup>3</sup> The double bonds are



**Figure 1.** Structures of pikromycin, amphotericin B, discodermolide, and curacin A.

formed by DH domains through abstraction of the  $\alpha$ -proton and concomitant protonation of the  $\beta$ -hydroxyl group of the nascent  $\beta$ -hydroxyacyl-ACP polyketide intermediate, resulting in loss of one water molecule. As observed in FAS DHs, the characteristic double hotdog fold is also found in PKS DHs to form the active site with two catalytic residues, aspartic acid and histidine.<sup>4</sup> The olefin geometry of DH products cannot be predicted through a signature fingerprint as in the KR<sup>5</sup> since the active site residues appear quite similar in both *cis*- and *trans*-olefin-generating DHs.<sup>4d,6</sup> Initial evidence suggested that the geometry of a double bond is exclusively dependent on the stereochemistry of the DH substrate provided by the upstream KR module, wherein A-type KR products (L-alcohols) lead to *cis*-olefins while *trans*-olefins arise from B-type KR products (D-alcohols);<sup>7</sup> however, exceptions have recently been observed.<sup>4e,8</sup> Post-PKS tailoring enzymes such as enoyl reductases or isomerases can further obscure the original olefin geometry.<sup>4e,8,9</sup>

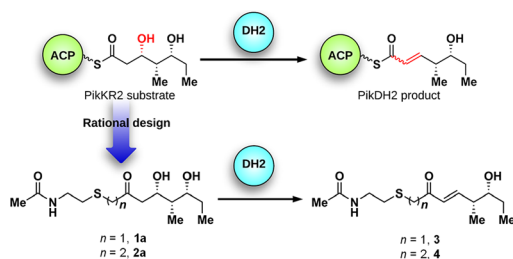
Steady-state kinetic analysis using simple diffusible N-acetylcysteamine (NAC) thioester precursors, which has been

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successfully employed to interrogate the substrate specificity of other PKS domains and modules,<sup>10</sup> has thus far been ineffective for studying isolated DH domains. This is likely due to the extremely low activity of the excised DH domains, which necessitates overnight incubations to generate sufficient product for detection. Moreover, the simple nature and ready reversibility of the dehydration reaction cannot be easily monitored by conventional radio-TLC or spectrophotometric assays. Cane and co-workers have also shown, at least in one example, that a NAC thioester substrate was not properly delivered to a DH active site, resulting in reversal of the diastereospecificity.<sup>4e</sup> This result highlights the potential importance of the ACP for chaperoning the polyketide intermediate for proper recognition and processing by DHs. Despite these aforementioned challenges, significant progress has been made in PKS functional characterization using NAC and ACP-bound substrates, which has provided useful insight into the substrate specificity and compelling evidence that (*E*)-unsaturated polyketide intermediates involve a stereospecific *syn* elimination.<sup>4e,8,11</sup> To further advance our understanding of DH domains, we report herein the first detailed steady-state kinetic characterization of an individual DH domain through monitoring of the reaction progress by liquid chromatography–tandem mass spectrometry (LC–MS/MS). The substrate specificity was studied with a systematic series of synthetic triketide analogues to probe the impact of both the vicinal and distal stereochemistry and through mutagenesis of active-site residues. The ability to monitor the reaction progress also permitted a study of the pH dependence of the catalytic activity and kinetic characterization of a mechanism-based irreversible inhibitor designed to facilitate cocrystallization and elucidation of substrate–protein interactions.

The pikromycin biosynthetic pathway is one of the most well-explored type I PKSs and has served as a model system for fundamental investigations of PKS catalysis.<sup>10d–f,12</sup> We selected the DH domain from pikromycin PKS module 2 (PikDH2) for our studies since it is responsible for the *trans*-olefin in pikromycin (Figure 1) and Cane and co-workers revealed that the cryptic  $\beta$ -hydroxyl stereochemistry of the triketide substrate possesses the *D*-configuration.<sup>10f,13</sup> We predicted that a larger multidomain portion of the PKS would create a more native context for *in vitro* analysis.<sup>13</sup> The region of *pikAI* encoding the PikKR2–DH2 didomain comprising residues 3579–4365 was cloned into a pMCSG7 expression vector, and the resulting protein was overexpressed and purified using standard methods (see the Supporting Information (SI)). Substrate mimics **1a** and **2a** for PikDH2 are based on its natural substrate (Figure 2).<sup>13</sup> While maintaining the same stereochemistry on the triketide moiety, they possess two major modifications compared with the natural substrate, including replacement of the ACP-phosphopantetheinyl arm with NAC and insertion of one or two



**Figure 2.** Rational design of PikDH2 substrate mimics **1a** and **2a** and the corresponding enzymatic products **3** and **4**.

methylene spacers between the carbonyl and the sulfur atom to avoid undesired intramolecular lactonization while not significantly perturbing the  $pK_a$  of the  $\alpha$ -protons.<sup>14</sup> These two modifications were tolerated by PikKR2, instilling confidence that the downstream DH would accept these substrates.<sup>13</sup> Overnight incubation of substrates **1a** and **2a** with the PikKR2–DH2 didomain on a relatively large scale generated the corresponding enzymatic products **3** and **4** in slightly more than 50% isolated yield (Scheme S4 in the SI). The structures of **3** and **4** were unambiguously confirmed by NMR spectroscopy and exhibited diagnostic <sup>13</sup>C chemical shifts at 150 and 130 ppm and a <sup>3</sup>J<sub>HH</sub> coupling of 16 Hz for the vinyl protons. These results are consistent with the empirical rule that *D*-alcohols provide *trans*-olefins.

Additionally, we investigated the reason for incomplete reaction. Since most FAS DHs conduct the reverse hydration reactions, enones **3** and **4** were incubated with PikKR2–DH2 to test the reversibility of the enzymatic reaction. We observed that PikKR2–DH2 stereoselectively converted **3** and **4** exclusively to their hydrated products **1a** and **2a**, respectively, whose identities were confirmed by LC–MS/MS with authentic synthetic standards (Figures S2 and S3 in the SI). When equilibrium was reached, the ratio of **1a** to **3** was 1:1.2 and the ratio of **2a** to **4** was 1:2, slightly favoring the dehydration products in both cases. In the biosynthetic pathway, this equilibrium is pushed toward dehydration, driven by downstream module activities.

To interrogate the stereospecificity of PikDH2, we altered the  $\beta$ -stereocenters of substrates **1a** and **2a** as well as every stereogenic center in **2a** through chemical synthesis (Schemes S1–S3), resulting in substrate stereoisomers **1b** and **2b–e** (Table 1).<sup>13</sup> As LC–MS/MS showed high sensitivity and

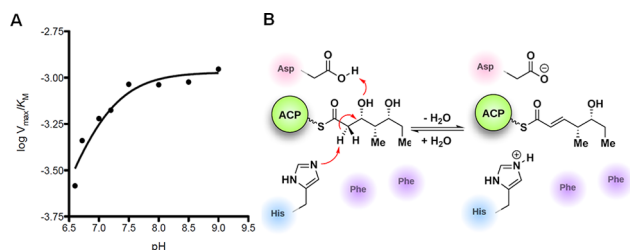
**Table 1.** Structures of Substrate Analogues **1a**, **1b**, **2a–e**, and **5** and Their Steady-State Kinetic Parameters

No.	Substrate Structure	$K_M$ (mM)	$k_{cat}$ ( $\text{min}^{-1}$ )	$k_{cat}/K_M$ ( $\text{min}^{-1}\text{M}^{-1}$ )
<b>1a</b>		$6.9 \pm 1.7$	$0.67 \pm 0.11$	$98 \pm 39$
<b>1b</b>		$> 8$	$< 0.01$	$< 1$
<b>2a</b>		$5.7 \pm 1.4$	$1.28 \pm 0.19$	$225 \pm 88$
<b>2b</b>		$> 8$	$< 0.01$	$< 1$
<b>2c</b>		$> 8$	$< 0.01$	$< 1$
<b>2d</b>		$> 8$	$< 0.01$	$< 1$
<b>2e</b>		$> 8$	$< 0.01$	$< 1$
<b>(±)-5</b>		$> 8$	$< 0.01$	$< 1$

selectivity for the detection of KR products,<sup>13</sup> we continued using this technique to examine the formation of DH products by their unique fragmentation patterns and/or retention times. The initial velocity was linear up to 40 min with  $10 \mu\text{M}$  PikKR2–DH2 didomain (Figure S4), and thus, we chose an end-point quench after 15 min of incubation for our kinetic studies. The kinetic

parameters for all of the substrates were obtained by fitting the saturation curves to the Michaelis–Menten equation (Table 1). Substrates **1a** and **2a** with the native stereochemistry were accepted and processed rapidly by the enzyme. The values of  $K_M$  ( $6.9 \pm 1.7$  and  $5.7 \pm 1.4$  mM) and  $k_{cat}$  ( $0.67 \pm 0.11$  and  $1.28 \pm 0.19$  min<sup>-1</sup>) were comparable, suggesting that the methylene spacer does not adversely impact active-site binding and catalysis. Substrates **1b** and **2b**, which are epimeric at the  $\beta$ -position, were not processed. This strict substrate specificity has been similarly observed with other DHs.<sup>4e,8a,11b</sup> To inspect the sensitivity of PikDH2 to changes in distal stereochemistry, we next investigated substrates **2c** and **2d**, which are epimeric at the  $\gamma$ - and  $\delta$ -positions, respectively, as well as **2e**, wherein both the  $\gamma$ - and  $\delta$ -positions are inverted. Enzymatic products of **2c–e** were not detected by LC–MS/MS, indicating an unprecedented degree of discrimination of these distal stereocenters. Moreover, racemic diketide **5**, similar to widely used diketide substrates to study other DHs,<sup>4e,8a</sup> was unexpectedly not converted to the corresponding dehydration product.

To study the chemical mechanism of PikDH2, the pH dependence of the catalytic efficiency ( $V_{max}/K_M$ ) with substrate **2a** was obtained from pH 6.6 to 9.0. The hyperbolic curve implicates at least one key ionizable group responsible for binding and catalysis (from either the free enzyme or the free substrate)<sup>15</sup> with a  $pK_a$  value of  $7.0 \pm 0.1$  (Figure 3A).



**Figure 3.** (A) pH dependence of  $\log(V_{max}/K_M)$ . (B) Mechanism and key residues involved in the dehydration reaction.

Protonation of this ionizable group at low pH abolished the enzyme activity. The  $pK_a$  values for the two hydroxyl groups of substrate **2a** are expected to be  $\sim 14$ – $16$ . Thus, we expect that the observed ionizable group is a general base in the enzyme, presumably the conserved histidine residue (His3611), which abstracts the  $\alpha$ -proton. An aspartic acid residue protonates the  $\beta$ -hydroxyl group to facilitate the loss of a water molecule (Figure 3B).<sup>4</sup> The slightly perturbed  $pK_a$  of 7.0 for the catalytic histidine, relative to the  $pK_a$  of 6.0 for the free amino acid, indicates the influence of adjacent residues to form a slightly negatively charged cavity.<sup>16</sup>

In order to understand the basis for the remarkable selectivity of PikDH2, we next aligned the amino acid sequence of PikDH2 with those of other DHs of known 3D structure.<sup>4c–e</sup> Two catalytic residues, the histidine (His3611) in an HXXGXXXP motif and the aspartic acid (Asp3800) in an HPALLD motif, are also found in PikDH2. However, the tyrosine in a YGP motif (observed in most DHs), proposed to assist the protonation of the  $\beta$ -hydroxyl group,<sup>6</sup> is absent in this DH. Instead, a phenylalanine (Phe3746, FGP motif) is located only 3.8 Å from the catalytic histidine in a PikDH2 homology model (Figure S7), suggesting that this residue could impact substrate binding at the active site. The F3746L point mutant was generated and tested with substrate surrogate **2a** to verify this possibility. The specificity constant dropped to 1/40 of that for

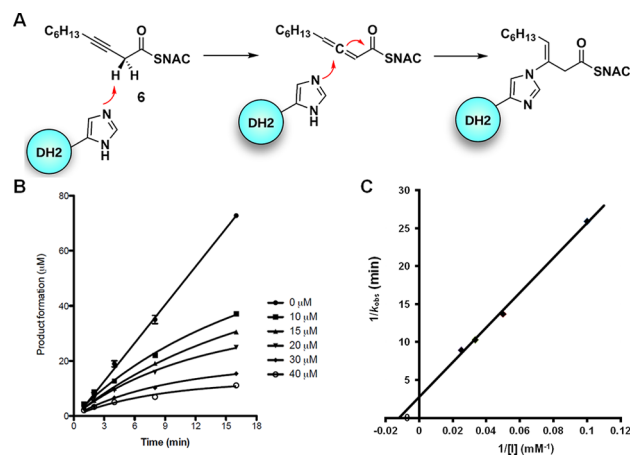
the wild type (WT), indicating the important role that this phenylalanine plays in substrate binding (Table 2). Another

**Table 2. Kinetic Parameters of PikDH2 Mutants**

mutant	$K_M$ (mM)	$k_{cat}$ (min <sup>-1</sup> )	$k_{cat}/K_M$ (min <sup>-1</sup> M <sup>-1</sup> )
WT	$5.7 \pm 1.4$	$1.28 \pm 0.19$	$225 \pm 88$
F3746L	>40	N/A	$5.2 \pm 0.3$
F3750Y	$4.1 \pm 1.4$	$0.086 \pm 0.013$	$21 \pm 9$
F3750L	$7.1 \pm 2.6$	$0.046 \pm 0.011$	$6.5 \pm 3.9$

phenylalanine, residue 3750, which is conserved in most DHs, is also near the catalytic residues (3.2 Å from the aspartic acid) in the homology model. The mutants F3750Y and F3750L had  $K_M$  values similar to that for the WT but 15–30-fold decreased  $k_{cat}$  values, demonstrating changing Phe3750 hampers the catalytic activity of PikDH2. We hypothesize that the two phenylalanine residues (F3746 and F3750) may shape a hydrophobic pocket at the active site to position the substrate as well as maintain the catalytically active state.

Cocrystallography of PKSs is particularly challenging since the substrate affinity is often low because of the intramolecular nature of substrate presentation. With the hope of aiding structural studies and potentially allowing visualization of protein interfaces between ACP and DH domains,<sup>17</sup> we set out to screen for a suitable PikDH2 suicide substrate that would covalently react within the enzyme active site. 3-Decynoyl-*N*-acetylcysteamine (**6**) developed by Bloch was the first mechanism-based inhibitor for any enzyme, which coincidentally inactivates FAS DHs.<sup>18</sup> Burkart and co-workers recently developed second-generation DH probes with improved chemical stability and demonstrated that they could inactivate functionally and structurally related PKS DHs.<sup>17</sup> Therefore, we evaluated the ability of **6** to irreversibly inhibit PikDH2 using our discontinuous assay monitoring the dehydration of substrate **2a** by LC–MS/MS. Compound **6** was shown to exhibit time-dependent inhibition, and the kinetic constants for inactivation were determined using the Kitz–Wilson method.<sup>19</sup> The kinetic parameters were  $K_I = 156 \pm 34$   $\mu$ M and  $k_{inact} = 0.36 \pm 0.06$  min<sup>-1</sup> (Figure 4B,C). The enzyme activity could not be recovered by dialysis, and substrate protected PikDH2 from inactivation.



**Figure 4.** (A) Mechanism of DH domain inactivation by 3-decynoyl-*N*-acetylcysteamine (**6**). (B) Time course of the inactivation of PikDH2 with 10–40  $\mu$ M **6**. Symbols represent the mean  $\pm$  SD from duplicate experiments. (C) Kitz–Wilson plot<sup>19</sup> of the inhibition data ( $1/k_{obs}$  vs  $1/[I]$ ).



Consequently, we expect that **6** covalently labels PikDH2 in analogy to FAS DHs through abstraction of an  $\alpha$ -proton by the catalytic histidine residue of the enzyme to form a reactive allene via propargylic rearrangement, which subsequently undergoes nucleophilic attack by the catalytic histidine (Figure 4A).<sup>20</sup>

In summary, we have performed the first steady-state kinetic analysis of a PKS DH domain by using LC–MS/MS analysis for product quantitation. PikDH2 was found to process only D-alcohols to furnish *trans*-olefin products. The enzyme showed an unforeseen extremely high stereospecificity at the  $\beta$ -,  $\gamma$ -, and  $\delta$ -positions of the substrate, potentially revealing a challenge for future metabolic engineering efforts. The pH dependence of  $k_{\text{cat}}/K_M$  identified a general base with a  $\text{p}K_a$  of 7.0 that is critical for substrate binding and catalysis, which we assign as a histidine (His3611). Mutagenesis and kinetic studies uncovered a critical hydrophobic region shaped by two phenylalanine residues that is essential for substrate binding as well as catalytic activity and suggested that the tyrosine in a YGP motif (observed in most DHs), which has been implicated in DH catalysis, is not required for activity. In the search for a cocrystallography candidate for PikDH2, we demonstrated that an alkyne-based probe irreversibly modified this noniterative type I PKS DH. These studies provide a foundation for future efforts aimed at uncovering the protein–substrate interactions that govern binding and catalysis. Our findings further reinforce the utility of diffusible NAC substrate mimics containing a ketone isostere in place of the labile thioester for studying intrinsically unstable PKS chain intermediates.

## ■ ASSOCIATED CONTENT

### ● Supporting Information

Experimental details and additional data. The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/jacs.5b02325.

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### Notes

The authors declare no competing financial interest.

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## ■ REFERENCES

- (1) (a) Smith, S.; Tsai, S.-C. *Nat. Prod. Rep.* **2007**, *24*, 1041. (b) Smith, J. L.; Sherman, D. H. *Science* **2008**, *321*, 1304.
- (2) (a) McDaniel, R.; Thamchaipenet, A.; Gustafsson, C.; Fu, H.; Betlach, M.; Ashley, G. *Proc. Natl. Acad. Sci. U.S.A.* **1999**, *96*, 1846. (b) Tang, L.; Shah, S.; Chung, L.; Camey, J.; Katz, L.; Khosla, C.; Julien, B. *Science* **2000**, *287*, 640. (c) Werneburg, M.; Busch, B.; He, J.; Richter, M. E.; Xiang, L.; Moore, B. S.; Roth, M.; Dahse, H. M.; Hertweck, C. *J. Am. Chem. Soc.* **2010**, *132*, 10407.
- (3) (a) Verdier-Pinard, P.; Lai, J.-Y.; Yoo, H.-D.; Yu, J.; Marquez, B.; Nagle, D. G.; Nambu, M.; White, J. D.; Falck, J. R.; Gerwick, W. H.; Day, B. W.; Hamel, E. *Mol. Pharmacol.* **1998**, *53*, 62. (b) Martello, L. A.; LaMarche, M. J.; He, L.; Beauchamp, T. J.; Smith, A. B., III; Horwitz, S. B. *Chem. Biol.* **2001**, *8*, 843.

- (4) (a) Leesong, M.; Henderson, B. S.; Gillig, J. R.; Schwab, J. M.; Smith, J. L. *Structure* **1996**, *4*, 253. (b) Kimber, M. S.; Martin, F.; Lu, Y.; Houston, S.; Vedadi, M.; Dharamsi, A.; Fiebig, K. M.; Schmid, M.; Rock, C. O. *J. Biol. Chem.* **2004**, *279*, 52593. (c) Keatinge-Clay, A. *J. Mol. Biol.* **2008**, *384*, 941. (d) Akey, D. L.; Razelun, J. R.; Tehranisa, J.; Sherman, D. H.; Gerwick, W. H.; Smith, J. L. *Structure* **2010**, *18*, 94. (e) Gay, D.; You, Y.-O.; Keatinge-Clay, A.; Cane, D. E. *Biochemistry* **2013**, *52*, 8916. (5) Caffrey, P. *ChemBioChem* **2003**, *4*, 654.
- (6) Keatinge-Clay, A. T. *Nat. Prod. Rep.* **2012**, *29*, 1050.
- (7) Reid, R.; Piagentini, M.; Rodriguez, E.; Ashley, G.; Viswanathan, N.; Carney, J.; Santi, D. V.; Hutchinson, C. R.; McDaniel, R. *Biochemistry* **2003**, *42*, 72.
- (8) (a) Vergnolle, O.; Hahn, F.; Baerga-Ortiz, A.; Leadlay, P. F.; Andexer, J. N. *ChemBioChem* **2011**, *12*, 1011. (b) Kandziora, N.; Andexer, J. N.; Moss, S. J.; Wilkinson, B.; Leadlay, P. F.; Hahn, F. *Chem. Sci.* **2014**, *5*, 3563.
- (9) Kusebauch, B.; Busch, B.; Scherlach, K.; Roth, M.; Hertweck, C. *Angew. Chem., Int. Ed.* **2010**, *49*, 1460.
- (10) (a) Holzbaur, I. E.; Harris, R. C.; Bycroft, M.; Cortes, J.; Bisang, C.; Staunton, J.; Rudd, B. A.; Leadlay, P. F. *Chem. Biol.* **1999**, *6*, 189. (b) Wu, N.; Kudo, F.; Cane, D. E.; Khosla, C. *J. Am. Chem. Soc.* **2000**, *122*, 4847. (c) Cane, D. E.; Kudo, F.; Kinoshita, K.; Khosla, C. *Chem. Biol.* **2002**, *9*, 131. (d) Lu, H.; Tsai, S.-C.; Khosla, C.; Cane, D. E. *Biochemistry* **2002**, *41*, 12590. (e) Yin, Y.; Lu, H.; Khosla, C.; Cane, D. E. *J. Am. Chem. Soc.* **2003**, *125*, 5671. (f) Wu, J.; Zaleski, T. J.; Valenzano, C.; Khosla, C.; Cane, D. E. *J. Am. Chem. Soc.* **2005**, *127*, 17393.
- (11) (a) Guo, X.; Liu, T.; Valenzano, C. R.; Deng, Z.; Cane, D. E. *J. Am. Chem. Soc.* **2010**, *132*, 14694. (b) Valenzano, C. R.; You, Y. O.; Garg, A.; Keatinge-Clay, A.; Khosla, C.; Cane, D. E. *J. Am. Chem. Soc.* **2010**, *132*, 14697. (c) Berkhan, G.; Hahn, F. *Angew. Chem., Int. Ed.* **2014**, *53*, 14240.
- (12) (a) Xue, Y.; Zhao, Li.; Liu, H.-W.; Sherman, D. H. *Proc. Natl. Acad. Sci. U.S.A.* **1998**, *95*, 12111. (b) Beck, B. J.; Aldrich, C. C.; Fecik, R. A.; Reynolds, K. A.; Sherman, D. A. *J. Am. Chem. Soc.* **2003**, *125*, 12551. (c) Kittendorf, J. D.; Sherman, D. H. *Bioorg. Med. Chem.* **2009**, *17*, 2137. (d) Dutta, S.; Whicher, J. R.; Hansen, D. A.; Hale, W. A.; Chemler, J. A.; Congdon, G. R.; Narayan, A. R.; Håkansson, K.; Sherman, D. H.; Smith, J. L.; Skiniotis, G. *Nature* **2014**, *510*, 512.
- (13) Li, Y.; Fiers, W. D.; Bernard, S. M.; Smith, J. L.; Aldrich, C. C.; Fecik, R. A. *ACS Chem. Biol.* **2014**, *9*, 2914.
- (14) The  $\text{p}K_a$  values of the  $\alpha$ -protons of thioesters and ketones are approximately the same because of the poor orbital overlap between the sulfur lone pairs and the carbonyl  $\pi^*$  orbital. For example, ethyl thioacetate has a  $\text{p}K_a$  of 21, whereas that of ethyl acetate is 26.5 and that of acetone is 20. See: Aymes, T. L.; Richard, J. P. *J. Am. Chem. Soc.* **1992**, *114*, 10297.
- (15) Fersht, A. *Structure and Mechanism in Protein Science: A Guide to Enzyme Catalysis and Protein Folding*; W. H. Freeman: New York, 1999; p 175.
- (16) Purich, D. L.; Allison, R. D. *Handbook of Biochemical Kinetics: A Guide to Dynamic Processes in the Molecular Life Sciences*; Academic Press: San Diego, 1999; p 405.
- (17) (a) Ishikawa, F.; Haushalter, R. W.; Burkart, M. D. *J. Am. Chem. Soc.* **2012**, *134*, 769. (b) Ishikawa, F.; Haushalter, R. W.; Lee, D. J.; Finzel, K.; Burkart, M. D. *J. Am. Chem. Soc.* **2013**, *135*, 8846.
- (18) (a) Brock, D. J.; Kass, L. R.; Bloch, K. J. *Biol. Chem.* **1967**, *242*, 4432. (b) Kass, L. R.; Bloch, K. *Proc. Natl. Acad. Sci. U.S.A.* **1967**, *58*, 1168.
- (19) Kitz, R.; Wilson, I. B. *J. Biol. Chem.* **1962**, *237*, 3245.
- (20) (a) Endo, K.; Helmkamp, G. M., Jr.; Bloch, K. *J. Biol. Chem.* **1970**, *245*, 4293. (b) Schwab, J. M.; Li, W. B.; Ho, C. K.; Townsend, C. A.; Salituro, G. M. *J. Am. Chem. Soc.* **1984**, *106*, 7293.